

Journal of Advances in Biology & Biotechnology

Volume 27, Issue 9, Page 296-303, 2024; Article no.JABB.121373 ISSN: 2394-1081

Impact of Iba and Naa Concentrations on Growth Characteristics of MR2 Mulberry (*Morus sinensis*) Using Mini-Clonal Technology at Nursery Level

Karthick Mani Bharathi B^a, Susikaran S.^{b*}, Parthiban KT^c, Vasanth V^a, Mohammad Rafiq Bhat^a, Ramya Harika K^a, Navaneetha Krishnan S^c, Sabarish M^a, Mithilasri. M^d, Kalpana R^a and Ashwin Niranjan M^c

 ^a Department of Sericulture, Forest College and Research Institute, Tamil Nadu Agricultural University, Mettupalayam, India.
^b Directorate of Open and Distance Learning, Tamil Nadu Agricultural University, Coimbatore, India.
^c Forest College and Research Institute, Tamil Nadu Agricultural University, Mettupalayam, India.

^d Center for Climate Change and Disaster Management, Anna University, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: https://doi.org/10.9734/jabb/2024/v27i91299

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: https://www.sdiarticle5.com/review-history/121373

Original Research Article

Received: 19/06/2024 Accepted: 21/08/2024 Published: 28/08/2024

ABSTRACT

Using mini-clonal technology, an experiment was carried out to examine the effects of rooting hormone on the growth and rooting characteristics of the mulberry (*Morus sinensis* L.) variety 'MR2'. Cuttings from the apical shoots of the mulberry variety MR2, which is the popular variety

*Corresponding author: E-mail: susi.agri@gmail.com;

Cite as: B, Karthick Mani Bharathi, Susikaran S., Parthiban KT, Vasanth V, Mohammad Rafiq Bhat, Ramya Harika K, Navaneetha Krishnan S, Sabarish M, Mithilasri. M, Kalpana R, and Ashwin Niranjan M. 2024. "Impact of Iba and Naa Concentrations on Growth Characteristics of MR2 Mulberry (Morus Sinensis) Using Mini-Clonal Technology at Nursery Level". Journal of Advances in Biology & Biotechnology 27 (9):296-303. https://doi.org/10.9734/jabb/2024/v27i91299.

among farmers and have good resistance towards mildew attack was taken for the study. The experiment was conducted at the Department of Sericulture, Forest College and Research Institute, Mettupalayam. Four replications of the Completely Randomized Design (CRD) experiment were used to arrange the data. Two hormones, such as Indole-3-butyric acid (IBA) and Napthalene acetic acid (NAA), were used in the experiment. The hormones were replicated four times in each of the following concentration ranges: 1000, 2000, 3000, 4000, and 5000 ppm. Different hormone concentrations were applied to apical shoot cuttings before they were planted in the appropriate rooting medium in a greenhouse. At the 90th day, measurements were taken of the following parameters like shoot length (cm), number of roots per plant (no's), root length (cm) and number of leaves per plant (no's). IBA and NAA at 5000 ppm out performed all other treatments in terms of mulberry mini cutting growth and root characteristics.

Keywords: Auxins; apical cuttings; biochemical; mulberry; mini clonal technology; MR2 variety.

1. INTRODUCTION

The Mulberry (Morus indica) is a member of the family Moraceae. It is a woody, deciduous perennial tree with a deeper root system that grows guickly [1]. There are roughly 68 mulberry species in the world; in India, the most often utilized species are *M. indica*, also known as the Indian mulberry, followed by *M. alba*, also known as the White mulberry, M. serrata, and M. laevigata [2]. With 28 chromosomes, the majority of the species in the genus Morus are diploid [3]. Mulberry grows between 28°N and 55°N latitude and can withstand a variety of climates, from temperate to tropical [4]. The nominal temperature range is 24°C to 26°C, with 600nominal 2500 mm of annual rainfall [5]. It has been grown all year round in the southern regions of India. specifically Tamil Nadu, Karnataka and Andra Pradesh [6].

Mulberry seeds have a 20-30% survival rate and germination, making seed replication low unfeasible [7]. Therefore, propagation techniques like as grafting and cutting are employed [8]. Mulberries are typically propagated in India using semi-hard wood cuttings, which are then either planted directly in the main field or grown in nurseries before being moved to the field [9]. Directly planting of cuttings in the main field frequently results in unfavourable development and a low survival rate [10]. Growing seasons and the cost of upkeep and care were the primary determinants of the rate of successful root growth of cuttings, even in nursery circumstances [11].

The auxin group of hormones is essential for the production of roots, and in nurseries, indole-3butyric acid (IBA) is a routinely employed hormone that is used in a variety of concentrations in conjunction with NAA [12]. By

accelerating the transition of rootina primordia and the transfer of sugars to the base of cuttings, they exhibit an indirect influence that resulted in the creation of active roots [13]. On the other hand, not much research has been done to assess how auxin affects the and growth development of mulberries. The goal of the current study was to clarify the impact of auxin on the rooting effectiveness of mulberry apical cuttings in this particular environment.

2. MATERIALS AND METHODS

Experiment conducted at 11°19'N, 76°56'E, 300 meters above sea level and 800 mm of rainfall, the experiment was conducted at the Department of Sericulture. Forest College and Research Institute, Mettupalayam. Cuttings from the apical shoots of the mulberry variety MR2, which is the popular variety among farmers and have good resistance towards mildew attack. Using sterile pruning secateurs, from the mother plants, these small cuttings were removed early in the morning to prevent excessive sun exposure [6]. Afterwards, they were cut with scissors to a length of 15 cm. Before being planted, a selection of cuttings were treated with a fungicidal solution (Carbendazim 50% WP) for 20 minutes, and then washed in distilled water to prevent desiccation [14]. IBA and NAA rooting hormones were made individually and at various concentrations in powder form viz., 1000, 2000, 3000, 4000, and 5000 parts per million. Talcbased formulations were made by combining talc powder, boric acid crystal, systemic fungicide and the necessary amount of rooting hormone (100 mg, 200 mg, 300 mg, 400 mg, and 500 mg/100 g of talc, respectively) with the remainder of stock solutions [15]. The treated cuttings were placed in a suitable rooting medium (Soil + FYM + Coir pith) and maintained under shade net in an inexpensive poly tunnel with a relative humidity of 75–80% and a temperature between 25 and 35°C [16]. They were also misted with water on occasion using rose cans and stored in a mist chamber. The shoot length, the number of roots per plant, the number of leaves per plant and the root length (cm) in each treatment were all observed after 90 days of planting in rooting mixture.

2.1 The Design of Statistics

Four replications of the Completely Randomized Design (CRD) experiment were used to arrange the data. According to the method developed by Panse and Sukhatme [17], data were subjected to Analysis of Variance (ANOVA) for significance and means were separated at the 95 per cent (P=0.05) significant level.

3. RESULTS AND DISCUSSION

3.1 Rooting Hormone's Impact on Mulberry Apical Cuttings Shoot Length

When comparing therapies with varying IBA and NAA concentrations, IBA at 5000 ppm showed the highest shoot length. At 5000 ppm, the shoot length was 22.20 cm in IBA, and 18.50 cm in NAA in MR2 variety. The administration of hormones stimulates and augments shoot development. According to Kalyoncu *et al.* [18], black mulberry cuttings with a IBA application had the maximum shoot length. Black mulberry cuttings treated with 5g I-1 IBA dose in bunch planting method suits for its superior shoot length, according to Koyuncu and Senel's [19].

According to Habibi [20], an increase in auxin concentrations caused the shoot length of oleander plants to grow up. A subsequent increase in IBA caused the shoot length to drop. In specific Triphlochiton scleroxylon clones, a larger dosage of auxin (200 µg per cutting) had been demonstrated to prevent roots and shoots growth in cuttings [21]. Likewise, in our investigation shoot length dropped below 3000 ppm of auxin concentration. Similar results have been reported by Husen *et al.* [22], Singh *et al.* [23], Singh *et al.* [24] in *Bougainvillea glabra*, Singh *et al.* [25] in Citrus lemon cv. Cuttings and Packialakshmi and Sudhagar [26] in teak mini cuttings, suggesting that the application of auxin could increase the shoot length in many species.

3.2 Rooting Hormone's Effect on Mulberry Apical Cutting's Root Number

The greatest number of roots was found in IBA compared to NAA when two hormones, such as IBA and NAA were used. At 90 DAP, IBA at 5000 ppm recorded the greatest number of roots (15.50 no's) followed by NAA at 5000 ppm (14.00 no's). In Psidium guajava, apical cuttings treated with IBA at 3000 ppm and high concentration developed more roots, which may have been caused by an optimal hormonal impact that accumulates necessary internal substances and promotes their downward movement [27] and Singh (2008). According to Ullah et al. [28], numerous species exhibit increased rooting in cuttings as a result of hormone treatment stimulating cambium activity. The fluctuation in dosage in relation to the quantity of roots might be led to the varietal and climatic variations in the location [29].

3.3 Effect of Rooting Hormone on Root Length of MR2 Apical Cuttings

The current study found that IBA recorded the highest root length across all treatments. The greatest root length measured at 90 DAP was 25.75 cm in IBA at 5000 ppm followed by 23.30 cm in IBA at 4000 ppm similarly 22.50 cm in NAA at 5000 ppm followed by 21.25 cm in NAA at 4000 ppm concentration. Similar hormonal effect was noted by Krishankumar [30] in mulberry leading to maximal root length. Maximum root length was observed by Ghatnatti in [31]. This was attributed to auxin activity, which may have been brought on by the hydrolysis and transport of carbohydrates towards the cuttings base, which resulted in cell division and elongation. According to Baroudi et al. [32], mulberry (Morus alba) softwood cuttings treated with 2000 ppm and high concentrations of IBA showed good root length, root number and rooting per cent. The results of Kumar [33] in Melia dubia and Galavi et al., [34] in Vitis vinifera are comparable to the current findings.

Treatments (IBA)	On 90DAP				
	Shoot length (cm) *	No of roots/plant (no's) *	Root length (cm) *	No of leaves/plant (no's) *	
IBA@1000ppm	12.50 ^e (3.67)	9.10 ^e (3.17)	15.20 ^e (4.02)	5.75 ^e (2.59)	
IBA@ 2000ppm	14.33 ^d (3.91)	10.75 ^d (3.42)	16.50 ^d (4.18)	6.70 ^d (2.77)	
IBA@3000ppm	16.50° (4.18)	12.00 ° (3.60)	20.33° (4.61)	7.50° (2.91)	
IBA@ 4000ppm	18.33 ^b (4.39)	13.33 ^b (3.78)	23.30 ^b (4.92)	9.25 ^b (3.20)	
IBA@ 5000ppm	22.20 ^ª (4.81)	15.50 ^a (4.06)	25.75 ^a (5.17)	11.33 ^a (3.5 [′] 1)	
Control	0.00 (1.00)	0.00 (1.00)	0.00 (1.00)	0.00 (1.00)	
SEd	0.04	0.05	0.07	0.07	
CD (.05%)	0.09*	0.13*	0.16*	0.15*	

Table 1. Effect of IBA on growth attributes of Morus sinensis

Significant @ P=0.05 level, Each value is the mean of four replications; () Values are square root transformed values

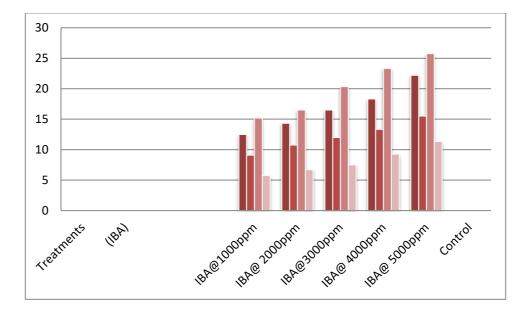


Fig. 1. Effect of IBA on growth attributes of Morus sinensis

Treatments (NAA)	On 90DAP				
	Shoot length (cm)*	No of roots/plant (no's) *	Root length (cm) *	No of leaves/plant (no's)	
NAA@ 1000ppm	11.25 ^e (3.50)	8.33 ^e (3.05)	14.25 ^e (3.90)	5.33 ^e (2.51)	
NAA@ 2000ppm	13.00 ^d (3.74)	10.25 ^d (3.35)	16.33 ^d (4.16)	6.00 ^d (2.64)	
NAA@ 3000ppm	16.33° (4.16)	11.00° (3.46)	19.10° (4.48)	7.22° (2.86)	
NAA@ 4000ppm	17.10 ^b (4.25)	12.33 ^b (3.65)	21.25 ^b (4.71)	8.22 ^b (3.03)	
NAA@ 5000ppm	18.50° (4.41)	14.00 ^a (3.87)	22.50ª (4.84)	10.50 ^a (3.39)	
Control	0.00 (1.00)	0.00 (1.00)	0.00 (1.00)	0.00 (1.00)	
SEd	0.06	0.05	0.08 `	0.05	
CD (.05%)	0.13*	0.12*	0.19*	0.10*	

Table 2. Effect of NAA on growth attributes of Morus sinensis

Significant @ P=0.05 level, Each value is the mean of four replications; () Values are square root transformed values

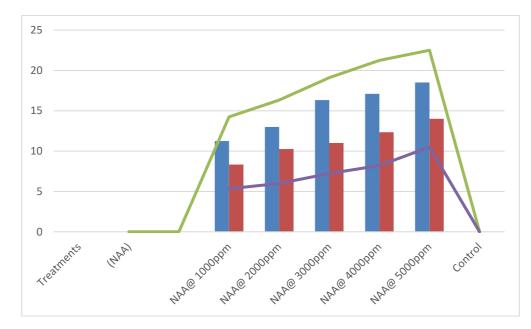


Fig. 2. Effect of NAA on growth attributes of Morus sinensis

3.4 Effect of Rooting Hormone on Number of Leaves of MR2 Apical Cuttings

In the current study, IBA at 5000 ppm recorded the most leaves of 11.33 no's at 90 DAP among the two treatments (IBA and NAA). IBA at 4000 ppm recorded the second-highest number of leaves of 9.25 no's. NAA at 5000 ppm registered the highest number of leaves of 10.50 no's. According to Pallavi et al. [29], there may be more roots, a higher plant, and more branches in mulberry cuttings at high ppm IBA concentration, which led to increase in leaves. According to Wahab et al. [35], there is a possibility that the influence of IBA on the number of leaves arises from the activation of shoot growth, which results in an increase in nodes and the subsequent formation of additional leaves in Psidium guajava L., Kiwi cuttings [36], Ficus, Hawaii [37] which supports the current study.

4. CONCLUSION

Using mini-clonal technology, the present investigation's findings indicate that IBA and NAA at 5000 ppm elicits the effective concentration for significant growth features in mulberry apical cuttings. Among all the treatment, IBA at 5000 ppm registered shoot length (22.20 cm), number of roots per plant (14.50 no's), root length (25.75 cm) and number of leaves per plant (11.33 cm). While NAA at 5000 ppm registered shoot length (18.50 cm), number of roots per plant (14.00 no's), root length (22.50 cm) and number of leaves per plant (10.50 cm). The main application of this mini-clonal technique is the multiplication of tree species. This is suggested due to its great rooting capability, ability to produce a larger number of plants annually, and affordable, high-quality root system. As a result, rooting hormone and its effective the concentration have been standardized in this work to enable mass mulberry proliferation in less time and space. Moreover, Mulberry is commonly propagated through cuttings but due to lack of 6 month old shoots for sapling production, mulberry sapling production becomes unstable but using mini clonal technique and standardization of rooting hormone, consistent mulberry sapling production is achievable and it is very helpful for the farmers.

DISCLAIMER (ARTIFICIAL INTELLIGENCE)

Author(s) hereby declare that NO generative AI technologies such as Large Language Models (ChatGPT, COPILOT, etc) and text-to-image

generators have been used during writing or editing of manuscripts.

ACKNOWLEDGEMENTS

There is no funding support for this work. All the authors acknowledge for the support provided by the Dean (FCRI), (Head & staffs) Deparment of Sericulture and Coordinator (Clonal complex).

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

- Bharathi B KM. A Comparative Biochemical Study of Mulberry (*Morus* spp.) Mini Clones Over Conventional Stem Cuttings. International Journal of Plant & Soil Science. 2024 Apr 13;36(5):975-83.
- 2. Datta RK. Mulberry cultivation and utilization in India. FAO Electronic Conference on Mulberry for Animal Production; 2000.
- 3. Sabarish M. Development of Mini clonal technology for Morus sinensis. M.Sc Thesis, Tamil Nadu Agricultural University, Coimbatore; 2017.
- 4. Singh A. A Note on Variation of Active Principles in Indian Medicinal Plants and TIM Formulations, Ethno botanical Leaflets. 2008;1:80.
- Hawramee OK, Aziz RR, Hassan DA. Propagation of white mulberry *Morus alba* L. fruitless cultivar using different cutting times and IBA. International Conference on AgriculturalSciences.2019;388.
- Karthick Mani Bharathi B, Susikaran S, Parthiban KT, Murugesh KA and K Chozhan. The economics of commercial mulberry saplings production using mini clonal technology over conventional method. The Pharma Innovation Journal. 2022; 11(7):1236-1241.
- Vijayan K, Ragunath MK, Das KK, Tikader A, Chakraborthi SP, Roy BN et al. Studies on leaf moisture of mulberry germplasm varieties. Indian J Seric. 1997; 36(2):155-157.
- 8. Hartmann HT, Kester D, Davies FT. Plant Propagation. Principles and Practices. Prentice Hall Inc., USA. Fifth edition, 1990.
- 9. Hartmann HT, Kester DE, Devies FT, Geneve RL. Plant Propagation: Principles

and Practices. 7th ed. Prentice-Hall. Englewood Cliffs, N.J, 2002.

- 10. Parthiban KT, Surendran C, Murugesh M, Buvaneswaran C. Vegetative propagation of a few multipurpose tree species using stem cuttings. Advances in Horticulture and Forestry, Jodhpur. 1999;6(27):175-178.
- Prakash D, Nivedha RM, Pushpadarini K, Ramazeame et al. Root and shoot growth of semi-hard wood cuttings of Mulberry (*Morus indica* L.) influenced by water imbibitions using wet cloth wrapping technique. International Journal of Scientific and Research Publications. 2017;7(5):2250-3153.
- 12. Ruppert DC. Hormone concentrations. Comb. Proc. Int. Plant Prop. Soc. 1974; 24:349-350.
- 13. Partiban KT, Seenivasan R. Forestry technologies A Complete value chain approach. Scientific Publishers, 2017. ISBN: 978-93-86102-60-7.
- 14. Kiruthika C, Susikaran S, Parthiban KT, Krishnamoorthy SV. Role of Auxins on growth of apical shoot cuttings of mulberry (*Morus indica* L.) using Mini clonal technology. IJCS. 2020;8(4):1896-9.
- Susikaran S, Shandeep SG, Haran MS, Deeikshana T, Abinaya C. Rooting Hormone and Substrate Effects On Mini-Cloned Mulberry (*Morus indica*). International Journal of Plant & Soil Science. 2023 Sep 16;35(20):72-83.
- Karthick Mani Bharathi B, Susikaran S, Parthiban KT, Vasanth V, Ashwin Niranjan M. Mass Production of Mulberry Saplings Generation Using Mini Clonal Technology: An Innovative Approach. 2024;5(4): 193-195.
- 17. Panse VG, Sukhatme. Statistical methods for agricultural workers. Indian Council of Agricultural Research, New Delhi, 1978.
- Kalyoncu IH, Ersoy N, Yýlmaz M, Aydýn M. Effects of humidity level and IBA dose application on the softwood top cuttings of white mulberry (*Morus alba* L.) and black mulberry (*Morus nigra* L.) types. African Journal of Biotechnology. 2009;8(16):3754-3760.
- 19. Koyuncu F, Senel E. Rooting of black mulberry (*Morus nigra* L.) hard wood cuttings, Journal of Fruit and Ornamental Plant Research. 2003;11:53-57.
- 20. Habibi KS. Effect of Auxin different concentrations on rooting of the semi hardwood cutting in oleander plant. Journal

of Plant Science Researchers. 2010;18(2):36-46.

- 21. Leakey RRB, Chapman VR, Longman KR. Physiological studies for tropical tree improvement and conservation. Factors affecting root initiation in cuttings of *Triplochiton scleroxylon* K. Schum. For. Ecol. Manage. 1982;4:53-66.
- Husen A, Muhammad Iqbal, Nasser S, Sohrab S, Masresha G. Effect of Indole-3-Butryic Acid on clonal propagation of Mulberry (*Morus alba* L.) stem cuttings: Rooting and associated biochemical changes, The National Academy of Sciences, India, 2015.
- Singh KK, Choudhary T, Kumar A. Effect of Various Concentrations of IBA and NAA on the Rooting of Stem Cuttings of Mulberry (*Morus alba* L.) under Mist House Condition in Garhwal Hill Region. Indian Journal of Hill Farming. 2014;27(1):125-131.
- Singh KK, Rawat JMS, Tomar YK. Influence of IBA on Rooting Potential of Torch Glory Bougainvillea glabra During Winter Season. Journal of Horticultural Science & Ornamental Plants 2011;3(2): 162-165.
- 25. Singh KK, Choudhary T, Kumar P. Effect of IBA concentrations on growth and rooting of Citrus limon cv. Pant Lemon cuttings. Hort Flora Research Spectrum. 2013;2(3):268-270.
- 26. Packialakshmi M, Jude sudhagar R. Standardization of rooting hormone in Mini clonal technology of Tectona grandis Linn. International Journal of chemical studies. 2019;7(3):4398-4401.
- Rani DT, Srihari D, Dorajeerao AVD, Subbaramamma P. Effect of rooting media and IBA treatments on root production and survival of terminal cuttings in guava (*Psidium guajava*) cv. Taiwan pink under mist house. International Journal of Chemical Studies. 2018;6(5): 2275-2281.
- 28. Ullah T, Wazir FU, Ahmad M, Analoui F, Khan MU, Ahmad M et al. A break through in guava (*Psidium guajava* L.) propagation from cutting. Asian Journal of Plant Sciences. 2005;4:238-43.
- 29. Pallavi D, Sharma GL, Naik EK. Effect of IBA and NAA on Rooting and Growth of Mulberry Cuttings. Int. J. Curr. Microbiol. App. Sci. 2018;7(11):305-308.
- 30. Krishan Kumar Singh. Effect of auxins and rooting media on rooting in stem cutting of

mulberry (*Morus nigra* L.). The Pharma Innovation Journal. 2018;7(11):12-15.

- Ghatnatti SA. Stuides on propagation of Duranta plumeri Jacq. Var. Goldiana by stem cuttings with growth regulators under mist. M.Sc. (Agri.) Thesis, University of Agricultural Sciences. Dharwad, 1997, 35-37.
- Baroudi H, Makhoul G, Mahfoud H. The effect of IBA (Indole-3-Butyric Acid) dose applications on rooting of hardwood and softwood top cuttings of white mulberry (*Morus alba* L.) and black mulberry (*Morus nigra* L.). Syrian Journal of Agricultural Research. 2017; 5(2): 23-34.
- Kumar P. Genetic evaluation, Growth characterization and clonal propagation studies in Melia dubia Cav. Ph.D. Thesis, Tamil Nadu Agricultural University, Coimbatore, 2011.

- Galavi M, Ali Karimian M, Mousavi SR. Effects of Different Auxin (IBA) Concentrations and Planting-Beds on Rooting Grape Cuttings (*Vitis vinifera*). Annual Review & Research in Biology. 2013; 3(4):517-523.
- Wahab F, Nabi G, Ali N, Shah M. Rooting response of semi hard wood cuttings of guava (*Psidium guajava* L.) to various concentrations of different auxins. Journal of Biological Sciences. 2001;1(4):184-87.
- Riaz A, Khalil UR, Muhammad I, Muhammad AR. Effect of indole butyric acid concentrations on the rooting of kiwi cuttings. Sarhad Journal of Agriculture. 2007; 23:293-95.
- Ismail SM, Asghar Hussain I. Effect of indole butyric acid and types of cuttings on root initiation of Ficus Hawaii. Sarhad Journal of Agriculture. 2007; 23(4): 919-925.

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of the publisher and/or the editor(s). This publisher and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

© Copyright (2024): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history: The peer review history for this paper can be accessed here: https://www.sdiarticle5.com/review-history/121373